

- activity has been proposed by A. Pullman, B. Pullman, and T. Nakajima, *Bull. Soc. Chim. Fr.*, 590 (1959).
- (6) G. P. Ceasar and J. J. Greene, *J. Med. Chem.*, **17**, 1122 (1974).
 - (7) J. F. Henderson and I. G. Junga, *Cancer Res.*, **21**, 118 (1961).
 - (8) R. J. Pugmire and D. M. Grant, *J. Am. Chem. Soc.*, **93**, 1880 (1971).
 - (9) R. J. Pugmire, D. M. Grant, L. B. Townsend, and R. K. Robins, *J. Am. Chem. Soc.*, **95**, 2791 (1973).
 - (10) M. T. Chenon, R. J. Pugmire, D. M. Grant, R. P. Panzica, and L. B. Townsend, *J. Heterocycl. Chem.*, **10**, 431 (1973).
 - (11) M. T. Chenon, R. P. Panzica, J. C. Smith, R. J. Pugmire, D. M. Grant, and L. B. Townsend, *J. Am. Chem. Soc.*, **98**, 4736 (1976).
 - (12) M. Dreyfus, G. Dodin, O. Bensaude, and J. E. Dubois, *J. Am. Chem. Soc.*, **97**, 2369 (1975).
 - (13) M. Dreyfus, O. Bensaude, G. Dodin, and J. E. Dubois, *J. Am. Chem. Soc.*, **98**, 6338 (1976).
 - (14) O. Bensaude, M. Dreyfus, G. Dodin, and J. E. Dubois, *J. Am. Chem. Soc.*, **99**, 4438 (1977).
 - (15) C. C. Cheng and R. K. Robins, *J. Org. Chem.*, **21**, 1240 (1956).
 - (16) E. C. Taylor and P. K. Loeffler, *J. Am. Chem. Soc.*, **82**, 3147 (1960).
 - (17) P. Schmidt, K. Eichenberger, M. Wilhelm, and J. Druey, *Helv. Chim. Acta*, **42**, 763 (1959).
 - (18) T. P. Johnston, A. L. Fikes, and J. A. Montgomery, *J. Org. Chem.*, **27**, 973 (1962).
 - (19) M. J. Cook, A. R. Katritzky, P. Linda, and R. D. Tack, *J. Chem. Soc., Perkin Trans. 2*, 1295 (1972).
 - (20) R. G. Bates, "Determination of pH", Wiley-Interscience, New York, N.Y., 1973, p 375.
 - (21) The same value of the enthalpy was obtained from the variation of the UV spectrum of 4APP with temperature. This is made possible by the absence of any extinction coefficient change with temperature (demonstrated by the absence of a fast relaxation phenomenon accompanying the chemical relaxation) at the wavelength of observation ($\lambda = 305$ nm), plus the fact that at this wavelength only the N(2)-H structure accounts for the overall absorbance of aqueous 4APP.
 - (22) When thermodynamically favorable, the protonation or OH⁻ fixation occurs at the same rate (i.e., that of H⁺ or OH⁻ diffusion) at the various basic or acidic sites (M. Eigen, *Angew. Chem., Int. Ed. Engl.*, **3**, 1 (1964)). This hypothesis is considered valid for all the kinetic studies presented in this work. Moreover, in order not to multiply the number of kinetic symbols, the same symbols (k_1 , k_{-1} , k_2 , k_{-2}) will be used for acid or base catalysis when not ambiguous.
 - (23) The protonation of the imidazole ring of adenine occurs at fairly low pH values as shown by the absence of any autocatalytic pathway for the tautomeric interconversion.¹² Assuming that the difference of basicity between pyrazole and imidazole rings in 4APP and adenine, respectively, is similar to the difference of basicity between pyrazole ($pK_a = 2.47$) and imidazole ($pK_a = 6.95$) leads to the reasonable conclusion that the common cation of 4APP would have a pK of formation around zero.
 - (24) R. A. Earl, R. J. Pugmire, G. R. Revankar, and L. B. Townsend, *J. Org. Chem.*, **40**, 1822 (1975).
 - (25) A very rough estimate of the proportion, α , of 7-H-1-*i*-Pr4APP cation is obtained by assuming that, in the absence of this tautomeric cation, the downfield shift at C_{7a} should be equal to the one at C₃ (-3.4 ppm) and that protonation at N₇ would produce a shift of 8.5 ppm at C_{7a}. This gives $-3.4(1 + \alpha) + 8.5\alpha = -1.9$; hence $\alpha \approx 10\%$.
 - (26) For difficulties related to the synthesis of the nontautomerizable model compounds of the various H tautomers, the attribution of the relaxation spectrum cannot be achieved by the usual comparison of the relaxation amplitude with the differential spectrum. The assignments will be tentatively made on the basis of other kinds of evidence provided by NMR or UV spectroscopy, as well as by the consistency of the hypothesis with the subsequent observations and conclusions.
 - (27) The precautions that must be taken when estimating the tautomeric equilibrium constants from the pKs of fixed derivatives have been reviewed by J. Elguero, C. Marzin, A. R. Katritzky, and P. Linda, *Adv. Heterocycl. Chem., Suppl. 1*, 15-29 (1976).
 - (28) (a) D. J. Brown and S. F. Mason, *J. Chem. Soc.*, 682 (1957); (b) S. F. Mason, *ibid.*, 1281 (1959); (c) *ibid.*, 3619 (1958); (d) J. J. Elliot and S. F. Mason, *ibid.*, 1275 (1959).
 - (29) B. C. Pal and C. A. Horton, *J. Chem. Soc.*, 400 (1964).
 - (30) M. Dreyfus, G. Dodin, O. Bensaude, and J. E. Dubois, *J. Am. Chem. Soc.*, **99**, 7027 (1977).
 - (31) The existence of an equilibrium between the imino structures 5-Me-1-H4IPP and 5-Me-2-H4IPP is far from being unlikely. Preliminary work carried out in this laboratory on 4-hydroxypyrazolo[3,4-*d*]pyrimidine (allopurinol), which is *isoelectronic* to 5-Me1PP, has established the presence of 1-H and 2-H tautomers in aqueous solutions. However, the attribution of the relaxation spectrum in neutral 5-Me4APP to the equilibrium involving the imino structures may be ruled out on the basis of the following arguments: (a) the relaxation amplitude should be much weaker than that actually observed, (b) the observed acidic autocatalytic cannot be understood since the cation common to both imino tautomers would not form abundantly when the imino forms are protonated.
 - (32) (a) M. Doree, personal communication; (b) M. Doree, P. Guerrier, and N. J. Leonard, *Proc. Natl. Acad. Sci. U.S.A.*, **73**, 1669 (1976).
 - (33) D. L. Nelson and G. B. Elion, *Biochem. Pharmacol.*, **24**, 1235 (1975).

Five-Membered-Ring Hydrogen Rearrangement in Mass Spectral Fragmentations. Another Mechanism of γ Cleavage

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Abstract: A new mechanism of γ cleavage in the mass spectra of aliphatic carbonyl compounds was characterized by examining the spectra of deuterium-labeled butanoic acids. γ cleavage occurs following five-membered-ring hydrogen transfer from the β carbon to an oxygen and the shift of a hydrogen atom from the α to the β carbon. This five-membered-ring hydrogen rearrangement is competitive with and occurs up to $1/3$ as frequently as the six-membered-ring hydrogen transfer which precedes the well known β -cleavage loss of an olefin.

The best known mass spectral rearrangement involves transfer of a γ hydrogen to a carbonyl oxygen in conjunction with olefin loss by β cleavage.²⁻⁴ Much evidence^{3,4} has indicated that six-membered-ring hydrogen transfer is highly specific, though competing hydrogen rearrangements by larger sized rings sometimes occur.⁴ We here demonstrate that five-membered-ring hydrogen rearrangement followed by γ cleavage³ competes significantly with six-membered-ring hydrogen transfer- β cleavage.

Results

Table I gives the intensities of the ions formed by γ cleavage and γ cleavage followed by the loss of a water molecule in the normal mass spectra of several deuterium-labeled butanoic acids and the intensities of the ions of the corresponding compositions in the spectra of 2-ethylbutanoic acid and 2-ethylbutanoic acid-*O*-*d*₁. The only detected metastable decompositions of the C₄H₈O₂⁺ ions were the losses of methyl

Table I. Abundances of Ions Formed by Losses of Methyl and of Methyl plus Water from Labeled Butanoic Acids

Compd	Isotopic composn	Ion abundances ^a							
		C ₃ H ₅ O ₂ ⁺	C ₃ H ₄ DO ₂ ⁺	C ₃ H ₃ -D ₂ O ₂ ⁺	C ₃ H ₂ -D ₃ O ₂ ⁺	C ₃ HD ₄ O ₂ ⁺	C ₃ H ₃ O ⁺	C ₃ H ₂ DO ⁺	C ₃ HD ₂ O ⁺
CH ₃ CH ₂ CH ₂ CO ₂ H		0.127					0.028		
CH ₃ CH ₂ CH ₂ CO ₂ D ^b	84% d ₁ , 16% d ₀	<0.010	0.112				0.032	0.003	
CH ₃ CH ₂ CD ₂ CO ₂ H ^{c,d}	97% d ₂ , 3% d ₁	0.014	0.004	0.100			0.001	0.003	0.018
CH ₃ CD ₂ CH ₂ CO ₂ H ^{c,e}	95% d ₂ , 5% d ₁	0.005	0.005	0.081			0.002	0.019	0.006
CD ₃ CH ₂ CH ₂ CO ₂ H ^{c,f}	96% d ₃ , 4% d ₂	0.12	0.012	0.004	0.008		0.027	<0.001	<0.001
CD ₃ CH ₂ CH ₂ CO ₂ D ^{b,f}	77% d ₄ , 22% d ₃	0.011	0.10	0.008	0.003	0.006	0.023	0.001	0.001
(CH ₃ CH ₂) ₂ CHCO ₂ H		0.164					0.101		
(CH ₃ CH ₂) ₂ CHCO ₂ D ^b	87% d ₁ , 13% d ₀	0.002	0.152				0.098	0.006	

^a All spectra were obtained at 70 eV at a source temperature of 210 °C on a Du Pont 21-491 mass spectrometer. All compounds were purified by gas chromatography. ^b Obtained by exchange with D₂O in the mass spectrometer inlet. ^c Corrected for a 0.015 contribution from the loss of OH. ^d Prepared from the Grignard reagent of 1-bromopropane-1,1-d₂, which was obtained by the reduction of propanoic anhydride to 1-propanol-1,1-d₂ with LiAlD₄ and conversion of the alcohol to 1-bromopropane-1,1-d₂ with HBr/H₂SO₄. ^e Prepared by the addition of bromoethane-2,2-d₂ to diethylmalonate. ^f Prepared by the addition of bromoethane-2,2,2-d₃ to diethylmalonate.

and ethylene, and the only metastable decomposition of C₃H₅O₂⁺ was the loss of H₂O. The intensities of the peaks resulting from those transitions are given in Table II. Loss of water followed by the loss of methyl from ionized butanoic acid probably does not contribute significantly to C₃H₃O⁺ formation, as the ion formed by the loss of H₂O is only 2% as abundant as that formed by the loss of methyl.

Discussion

γ-Hydrogen rearrangement-β cleavage of 2-ethylbutanoic acid gives ion **4** in Scheme I. Metastable decompositions (Table II) demonstrate that ionized butanoic acid and **4** both lose methyl. Loss of methyl from **4** gives **5** by γ cleavage, as enolic ions fragment in a parallel fashion.^{5,6} Methyl is also lost from butanoic acid by γ cleavage, as 86% of the methyls lost from butanoic acid-4,4,4-d₃ were ·CD₃. This implies that ionized butanoic acid rearranges to **4** and then loses methyl to form **5**.

Metastable decompositions (Table II) demonstrate that C₃H₅O₂⁺ formed from both butanoic acid and 2-ethylbutanoic acid fragments by loss of H₂O. Largely HDO was lost from 5-O-d₁ derived from 2-ethylbutanoic acid-O-d₁. Therefore, both hydrogens in the water lost from **5** must be on the oxygens, i.e., **5** must decompose by **5** → **6**, as at least 50% H₂O would

be lost from 5-O-d₁ by any other mechanism. Since C₃H₄DO₂⁺ formed by loss of methyl from butanoic acid-O-d₁ also lost mostly HDO, **5** must be formed by the loss of methyl from ionized butanoic acid. Thus, the butanoic acid ion must rearrange to **4** prior to losing methyl, as **4** is the only logical C₄H₈O₂⁺ precursor of **5**. HDO was also predominantly lost from **5** formed from butanoic acid-3,3-d₂, while dominantly D₂O was lost from **5** obtained from butanoic acid-3,3-d₂-O-d₁. This demonstrates that the second hydrogen on the oxygen of **5** generated from butanoic acid comes from the β carbon, and therefore the occurrence of **1** → **3**. Other mechanisms placing a β hydrogen in the water lost are very improbable. The rearrangement of a β hydrogen to oxygen in ·CH₂-CH₂C(=O⁺)OH formed by the loss of methyl directly from **1** or the concerted loss of H₂O containing a β hydrogen from that species would give energetically highly disfavored :CHCH₂C≡O⁺. It is also unlikely that the loss of water specifically containing a β hydrogen would occur from the cyclic ions which have been suggested³ as possible products of this γ cleavage. Thus, **1** → **3**, **4** → **5**, and **5** → **6** occur in sequence in the decomposition of ionized butanoic acid, so the predominant mechanism for the reported losses of methyl and water must be **1** → **3** → **4** → **5** → **6**.

5 is 22% as abundant as **7** in the normal spectrum of buta-

Scheme I

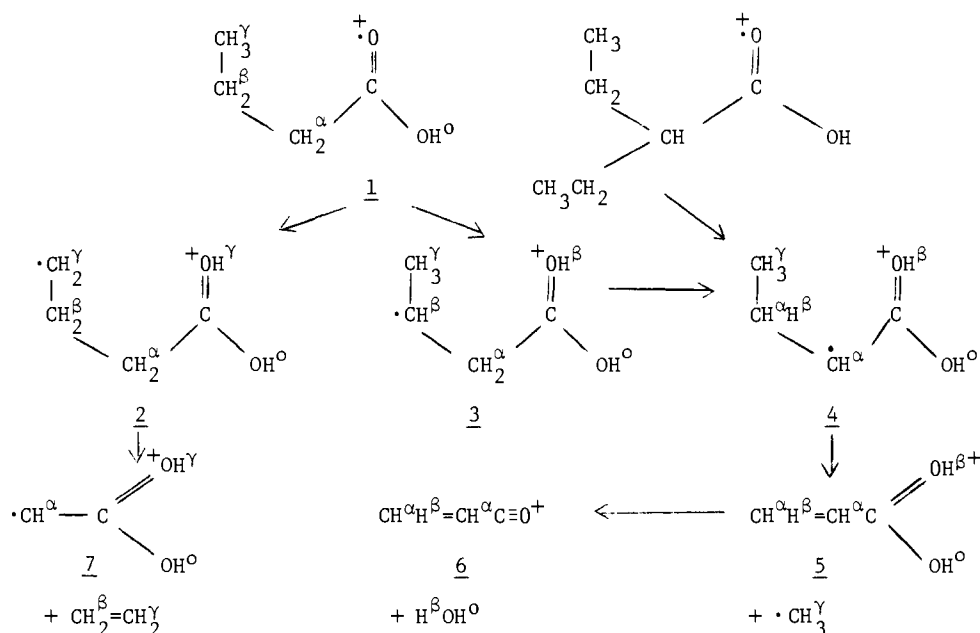


Table II. Metastable Decompositions of $C_4H_8O_2^+$ and $C_3H_5O_2^+$ ^a

Compd	Ion	-CH ₃	-C ₂ H ₄	-H ₂ O
CH ₃ CH ₂ CH ₂ CO ₂ H	1	2.4×10^{-2}	1.2×10^{-2}	
(CH ₃ CH ₂) ₂ CHCO ₂ H	4	9.6×10^{-3}	9.0×10^{-5}	
CH ₃ CH ₂ CH ₂ CO ₂ H	5			4.5×10^{-3}
(CH ₃ CH ₂) ₂ CHCO ₂ H	5			2.0×10^{-3}

^a Values are the intensities of the peaks representing the metastable transitions divided by the intensities of the peaks in the normal spectra representing the precursor ions. The intensities of the peaks representing metastable transitions were determined by metastable defocussing achieved by lowering the electrostatic analyzer potential at constant accelerating potential.

noic acid. The average of the ratios of the intensities of the ions produced by γ cleavage to those produced by γ -hydrogen rearrangement- β cleavage is ~ 0.3 in the mass spectra⁷ of acids, esters, and aldehydes with *n*-propyl and *n*-butyl moieties attached to their carbonyl groups. Therefore, five-membered-ring hydrogen rearrangements occur up to $1/3$ as frequently as competing six-membered-ring hydrogen rearrangements.

Hydrogen rearrangements via six-, seven-, and eight-membered rings followed by further rearrangement to ions

analogous to **5** also lead to γ cleavage.^{5,8} HDO was lost in 44% of the metastable decompositions of the $C_3H_3D_2O_2^+$ ions formed from hexanoic acid-3,3-*d*₂. The first step leading to the formation of the ions that lost Hdo must have been five-membered-ring hydrogen transfer to an oxygen. Since the γ -cleavage product $C_3H_3D_2O_2^+$ was 45% as abundant as $C_2H_4O_2^+$ formed by γ -hydrogen rearrangement- β cleavage in the spectrum of hexanoic acid-3,3-*d*₂, five-membered-ring hydrogen rearrangements probably generally accompany six-membered-ring rearrangements.

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References and Notes

- (1) (a) University of Texas Medical Branch; (b) Cornell University
- (2) F. W. McLafferty, *Anal. Chem.*, **31**, 82-87 (1959).
- (3) H. Budzikiewicz, C. Djerassi, and D. H. Williams, "Mass Spectrometry of Organic Compounds", Holden-Day, San Francisco, Calif., 1967, pp 138, 155-162.
- (4) D. G. I. Kingston, J. T. Bursley, and M. M. Bursley, *Chem. Rev.*, **74**, 215-242.
- (5) G. Eadon, *J. Am. Chem. Soc.*, **94**, 8938-8939 (1972).
- (6) W. Carpenter, A. M. Duffield, and C. Djerassi, *J. Am. Chem. Soc.*, **90**, 160-164 (1968).
- (7) Mass Spectral Data Collection, Mass Spectrometry Data Centre, A.W.R.E., Aldermaston, England.
- (8) R. Liedtke and C. Djerassi, *J. Am. Chem. Soc.*, **91**, 6814-6821 (1969).

Degenerate Rearrangements in Solvolytic Studies with *cis*- and *trans*-2-Phenyl-1,2-di-*p*-tolylvinyl-2-¹³C Bromides

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Abstract: Acetolysis in the presence of AgOAc of either *cis*- or *trans*-2-phenyl-1,2-di-*p*-tolylvinyl-2-¹³C bromide (*cis*- or *trans*-1-Br-2-¹³C) gave a 1:1 mixture of *cis* and *trans* products. After conversion of this product mixture to 2-phenyl-1,2-di-*p*-tolylethanol-*x*-¹³C (2-*x*-¹³C) and upon analysis of its ¹³C NMR spectrum, about the same extent (1.5-2.0%) of scrambling of the ¹³C label from C-2 to C-1 was found for either the *cis* or *trans* reactant. Nearly the same rate was also observed for the acetolysis, in the presence of NaOAc, with either *cis*- or *trans*-1-Br as substrate. Similarly, trifluoroacetolysis in the presence of CF₃COOAg of either *cis*- or *trans*-1-Br-2-¹³C also gave about the same extent of ¹³C scrambling (34-35%). All of these results point to the formation, without phenyl participation, of a free 2-phenyl-1,2-di-*p*-tolylvinyl cation which could then undergo competitively degenerate rearrangement by 1,2-phenyl shift and solvent capture to give product, the less nucleophilic the solvent, the greater the extent of isotopic scrambling. A solvent isotope effect, k_H/k_D , of 3.4-3.9 was observed for the reaction of *cis*- and *trans*-1-Br in CF₃COOH or CF₃COOD, without the presence of any Ag salt. This finding indicated that, in the reaction with CF₃COOH in the absence of Ag salt, an electrophilic addition-elimination process must have played an important role. Reaction of *cis*- and *trans*-1-Br-2-¹³C with CF₃COOH, without any CF₃COOAg, gave 45 and 48-49% scrambling after ~ 2.5 and 6 half-lives. It is suggested that these latter results may be chiefly attributable to a subsequent ionization, in the reaction medium, of the addition-elimination product, followed by degenerate 1,2-phenyl shifts and recombination with solvent.

Degenerate rearrangements from 1,2-aryl shifts across the double bond in a number of labeled triarylvinyli cations, with various combinations of phenyl and/or *p*-anisyl as the aryl groups, have been studied in this laboratory,¹ and by Rappoport and coworkers.² 1,2-Phenyl and 1,2-anisyl shifts in triphenylvinyl and trianisylvinyl cationic systems have been investigated using the ¹⁴C label as tracer,^{1a,b,e} and using ¹³C labeling coupled with analysis by ¹³C NMR,^{1b,c} while the ¹³C NMR technique has also been applied in a study on the possible 1,2-phenyl shift in the reaction of *cis*- and *trans*-1,2-

dianisyl-2-phenylvinyl-2-¹³C bromide with HOAc-AgOAc.^{1d} Rappoport et al. have utilized a D-labeled phenyl group and ¹H NMR as well as mass spectrometry in investigating degenerate 1,2-anisyl shifts during the solvolysis of *cis*- and *trans*-2-anisyl-1,2-diphenylvinyl bromides.^{2a} Degenerate rearrangements in the trianisylvinyl and the *cis*- and *trans*-1,2-dianisyl-2-phenylvinyl systems have also been studied by Rappoport et al. in a number of solvolytic reactions with a CD₃OC₆H₄ group as label and again with analyses by ¹H NMR and mass spectrometry.^{2b} Very recently, we have in-